

Review Article

$\mbox{PPAR}\delta$, a Potential Therapeutic Target for Heart Disease

Qinglin Yang¹ and Qinqiang Long²

¹Cardiovascular Center of Excellence, LSU Healther Science Center, 533 Bolivar St, New Orleans, LA 70112, USA

²Division of Cardiology, Department of Internal Medicine, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, 1095 Jiefang Ave, Wuhan, 430030, China

Abstract. The nuclear receptor peroxisome proliferator-activated receptor δ (PPAR δ) can transcriptionally regulate target genes. PPAR δ exerts essential regulatory functions in the heart, which requires constant energy supply. PPAR δ plays a key role in energy metabolism, controlling not only fatty acid (FA) and glucose oxidation, but also redox homeostasis, mitochondrial biogenesis, inflammation, and cardiomyocyte proliferation. PPAR δ signaling is impaired in the heart under various pathological conditions, such as pathological cardiac hypertrophy, myocardial ischemia/reperfusion, doxorubicin cardiotoxicity and diabetic cardiomyopathy. PPAR δ deficiency in the heart leads to cardiac dysfunction, myocardial lipid accumulation, cardiac hypertrophy/remodeling and heart failure. This article provides an up-today overview of this research area and discusses the role of PPAR δ in the heart in light of the complex mechanisms of its transcriptional regulation and its potential as a translatable therapeutic target for the treatment of cardiac disorders.

Keywords: PPARs, transcription regulation, cardiac hypertrophy, heart failure, cardiac disorders, fatty acid metabolism, mitochondrial biogenesis, inflammation, oxidative stress

1. Introduction

Peroxisome proliferator-activated receptors (PPAR α , - γ and - δ) belong to a nuclear receptor transcription factor superfamily that regulates metabolic transcription. Ligands of PPAR α and - γ are clinically effective in treating dyslipidemia and insulin resistance, although adverse risks have remained a concern (see review [1]). PPAR δ (NR1C2, encoded by the Ppard gene, also known as PPAR β) was first cloned in the early 90's of the 20th century [2, 3]. No FDA approved drug targeting PPAR δ is available so far notwithstanding early-stage clinical trials on the use of PPAR δ agonists (e.g., KD-3010 and MBX-8025) to treat atherosclerosis, metabolic disorders and liver injury [4–6]. Early-stage clinical trials of selective PPAR δ agonists Seladelpar (MBX-8025) in patients with primary biliary cholangitis was terminated due to grade 3 increases in serum transaminase levels [7]. Therefore, a PPAR δ -specific therapeutic strategy remains in perspective with improved selectivity and minimal off-target effects.

As the most energy demanding organ, energy metabolism is essential for the normal function of hearts. A normal adult heart predominantly relies on the energy from fatty acid oxidation (FAO). PPARs play essential roles in transcriptional regulation of many aspects of cellular metabolism, especially FAO. All three PPAR subtypes are expressed in heart cells

Qinglin Yang qyang1@lsuhsc.edu

Editor

Yves Lecarpentier

Dates

Received 20 June 2018 Accepted 3 October 2018

Copyright © 2018 Qinglin Yang and Qinqiang Long. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



(cardiomyocytes) [8–11]. PPAR α controls the transcriptional expression of key enzymes that are involved in FA uptake and oxidation, triglyceride synthesis, mitochondrial respiration uncoupling and glucose metabolism [10, 12, 13]. The role of PPAR γ in the heart has not been extensively studied due to its relatively low abundance. However, several studies on cardiomyocyte-restricted PPAR γ mice demonstrate its crucial role in cardiac physiology, probably via its role in transcriptionally regulating myocardial FA metabolism and anti-oxidant defense [14–17]. The cardiac function of PPAR δ been explored until subtype-specific synthetic ligands for PPAR δ became available in the early 2,000's. In the heart, PPAR δ is prominently expressed in cardiomyocytes and activation of PPAR δ upregulates FAO transcripts as in cultured cardiomyocytes [18, 19]. Global PPAR δ KO in mice is embryonically lethal due to placenta defects [20]. Survival PPAR δ KO mice show an extremely lean phenotype [20]. Considering that potential compensation may conceal potential phenotypic manifestation, conditional PPAR δ KO mouse lines were developed [20–29]. Inborn cardiomyocyte-restricted PPAR δ deletion leads to cardiomyopathy and heart failure with impaired myocardial FAO [30]. Therefore, it becomes clear that all PPAR subtypes play roles in transcriptional regulation of myocardial energy and lipid homeostasis, but with differences in their cardiac expression levels and yet to defined differential regulation on various aspects of energy metabolism. Further studies on the potential distinctive roles of each PPAR subtype in the heart should provide new therapeutic targets for treating heart disease. In this review, we focus on summarizing the biological function and clinical implications of PPAR δ in the heart.

2. PPAR& Plays a Crucial Role in Cardiac Pathophysiology

The optimal coupling of metabolism and function is essential for the normal structure/function of the heart. Cardiac metabolism is a key player in the development of cardiac pathophysiology as FAO provides a majority of energy for a normal heart to properly function [31, 32]. Metabolic adaptation becomes essential to meet the energy needs of the pathological heart ensue with various stimuli, such as mechanical overload, ischemia/reperfusion, aging, diabetes and drug cytotoxicity. Others and our studies documented that PPAR δ is a key transcription factor that regulates almost all steps of metabolism (see review [9, 33], hence PPAR δ can play a crucial role in determining cardiac pathophysiology.

Under various cardiac hypertrophy conditions, most studies show cardiac PPAR δ is decreased [34, 35] as those of PPAR α [12, 13]. PPAR δ is reduced in hypertrophic cardiomyocytes induced by angiotensin II [34] and hypertrophic hearts resulted from transverse aortic constriction (TAC)-induced left ventricular pressure overload [35]. The impaired cardiac PPAR δ expression could be reversed by anti-hypertrophic treatment [34, 36]. Studies using Western blots also showed protein contents of cardiac PPAR δ are downregulated in doxorubicin-cardiac cytotoxicity [37] and STZ-induced diabetic hearts [38]. In contrast to the above investigations, another study using Western blot showed that cardiac PPAR δ is upregulated in the heart with infarction in rats [39]. The reason for these different findings of PPAR δ in pathological hearts could be related to the specific pathological condition, the animal species/strain, and the technical issues of antibody specificity.

While changes of PPAR δ expression in the pathological heart need further validation, the essential role of cardiac PPAR δ is well established. Cardiomyocte-restricted PPAR δ deletion

was achieved by *breeding* the floxed mice with a mouse line with cardiomyocyte-restricted expression of Cre recombinase (α -Myosin heavy chain-Cre). Inborn cardiomyocyte-restricted PPAR δ deletion impairs myocardial FAO and leads to cardiomyopathy and heart failure [30]. To further gain insights into the function of PPAR δ in the adult heart, we generated mice with Tamoxifen-induced cardiomyocyte-restricted PPAR δ deletion by crossing the floxed PPAR δ mice with a mouse line carrying α -Myosin heavy chain-Mer-Cre-mer. Adult mice with cardiomyocyte-restricted PPAR δ deletion consistently show cardiac dysfunction, cardiac hypertrophy and remodeling [40]. On the other hand, studies using *gain-of-function* approaches also mostly support a protective effect of PPAR δ in the heart. Transgenic mice with cardiomyocyterestricted expression of a constitutively active form of PPAR δ show no overt cardiac phenotype, yet these mice become resistant to pressure overload-induced cardiac dysfunction [41].

Since the role and activity of PPAR δ is not entirely dependent on its transcript or protein abundance but also on the molecular regulation that PPARS exerts as gene inducer or repressor, studying the effects of pharmacological activators is therefore complementary to abundance studies. PPAR δ selective ligands reduce hypertrophic response induced by phenylephrine in cultured rat neonatal cardiomyocytes and rat embryonic myocytes (H9C2) [42]. PPAR δ activation improves angiotensin II-induced cardiac hypertrophy in vitro [43]. However, a later study on cultured rat adult cardiomyocytes shows upregulation of PPAR δ activity and FAO gene expression with minimal effect on cell growth [43]. It remains unclear whether the discrepancy is related to the differentiation states of cardiomyocytes or the culture conditions used in the studies. In vivo pharmacological studies in the literature provide further support. An early study on rats shows the PPAR δ agonist GW0742 protects against right heart hypertrophy from postmyocardial infarction [44]. Pharmacological studies show that baicalin and Moringa oleifera Seeds, both extracted from plants, protect the heart from TAC-induced pressure overload or hypertension from the development of hypertrophy via enhancing cardiac PPAR δ expression [35, 45]. The cardiac protective effect of PPAR δ is further supported by studies in animal models with myocardial ischemia/reperfusion (IR) injury. Activation of PPAR δ attenuates myocardial IR injury in rats [39] and mice [46]. Our results from a study on mice with cardiomyocyterestricted transgenic expression of the constitutively active PPAR δ supports the protective role of PPAR δ activation. Mice with PPAR δ activation showed improved cardiac function, reduced infarct size, and fibrotic remodeling [47]. Treatment with GW0742 protects the heart in a PPAR δ dependent manner [47]. It is interesting that another study on the effect of permanent myocardial infarct in rats shows functional effects of PPAR δ agonist (GW0742) treatment, but with augmented cardiac fibrosis with no functional consequece [48]. However, a control group to determine the basal effect of PPAR δ agonist (GW0742) was not included in this study, missing the opportunity to determine if GW0742 treatment alone could induce cardiac fibrosis. It is noted that the study assigned hearts to different groups based on the degree of infarction determined by echocardiographic images were used for all experimental procedures. The overtly biased experimental design may oblivion correct interpretation of the outcomes. Further studies with careful experimental design should help clarify the conclusions.

The cardiac protective role of PPAR δ activation has been validated further in other advert states. Cardiac cytotoxicity is a main clinical dilemma in the use of effective anti-cancer drugs. Evidence has emerged that activation of PPAR δ protects against doxorubicin-induced cardiotox-icity. A PPAR δ activator, L-165041, inhibits senescence in neonatal rat cardiomyocytes and

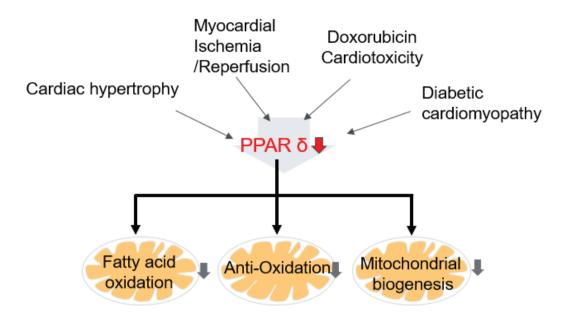


Figure 1: **PPAR** δ **plays a crucial role in cardiac pathophysiology:** PPAR δ signaling is impaired in the heart under various pathological conditions, such as pathological cardiac hypertrophy, myocardial ischemia/reperfusion, doxorubicin cardiotoxicity and diabetic cardiomyopathy. PPAR δ deficiency in the heart leads to cardiac dysfunction, myocardial lipid accumulation, cardiac hypertrophy/remodeling and heart failure.

H9C2 cells via interacting with Bcl6 to reduce apoptosis [49]. On the other hand, PPAR δ ligand GW0742 exerts a protective role in neonatal rat cardiomyocyte treated with Doxorubicin by sustaining intracellular calcium concentration [50]. Baicalein, a natural PPAR δ ligand, protects against doxorubicin-induced cardiotoxicity by attenuation of mitochondrial oxidant injury and JNK activation [51].

Diabetes (type 1 and type 2) and obesity have a strong link to cardiac dysfunction, cardiac hypertrophy and heart failure (see review [52–54]). Normalizing cardiac PPAR δ protein improves cardiac fibrosis in rats with streptozocin (STZ)-induced diabetes [37]. Induction of cardiac angiopoietin Like 4 (Angptl4) via PPAR δ activation in the heart is essential in protecting against FA-induced oxidative stress [55], a key mechanism behind diabetic cardiomyopathy. Several reports indicate that restoring PPAR δ protein expression in diabetic hearts from rats induced by STZ by various pharmacological compounds, such as histone deacetylase (HDAC) inhibitor [56], ginseng [9, 57] and ramipril [58]. However, many of these preliminary findings need further validation. Further studies are required to assess the PPAR δ expression in the heart in transcript level, target gene expression and subtype-specific activity to confirm the observations.

In summary, It is apparent that most of the current studies illustrate that cardiac PPAR δ may be repressed in cardiac disorders and PPAR δ re-activation in the heart is protective (Figure 1), whereas PPAR α re-activation in the heart could be detrimental [59, 60] or beneficial [61–64], possibly depending on the degree of activation.

3. Molecular and Biochemical Mechanisms Underlying the Biological Action of PPARδ in the Heart

3.1. PPARδ is a key transcriptional regulator of FA utilization in the heart

As a ligand-activated nuclear receptor and transcription factor, PPAR δ regulates transcription in many tissues (see review [9, 65, 66]). Being ubiquitously expressed in the body, PPAR δ is expressed preferentially in cardiomyocytes among other cell types in the heart [19]. In rat neonatal cardiomyocytes (RNCM), PPAR δ regulates transcript expression of key genes involved in FA metabolism [19]. PPAR δ -selective ligand and PPAR δ overexpression in cultured RNCM promotes FA metabolic gene expression and the rate of FAO in a classic ligand binding dependent mechanism [19]. We and other further confirmed that PPAR δ regulates the transcriptome of FA utilization in the heart (see reviews [9, 67, 68]). PPAR δ is a key determinant of FA uptake in cardiomyocytes by upregulating FA uptake genes, such as FA transport protein (FATP) and FA binding protein (FABP) [18, 19, 30, 40, 41, 69, 70]. Moreover, PPAR δ ligand treatment not only upregulates key mitochondria-specific β -oxidation genes, such as Longchain acyl-CoA synthetase (LACS), carnitine palmitoyltransferase very-long-chain acyl-CoA dehydrogenase (VLCAD), mitochondrial 3-ketoacyl-CoA thiolase, CPT-A, uncoupling protein 2 (UCP2), UCP3, pyruvate dehydrogenase kinase 4 (PDK4), medium chain acyl CoA dehydrogenase (MCAD), long chain acyl CoA dehydrogenase (LCAD) and malonyl CoA decarboxylase (MCD), but also peroxisome-specific β -oxidation genes, such as acyl-CoA (ACO), and peroxisomal 3-ketoacyl-CoA thiolase [18, 19, 30, 40, 41, 69]. Further studies on cardiacspecific PPAR δ mice provide definitive evidence supporting the role of PPAR δ in maintains constitutive FA metabolism in the heart [19]. In addition to depressed FAO, cardiomyocyterestricted PPAR δ knockout mice develop severe phenotypic changes, such as cardiac dysfunction, myocardial lipid accumulation and progressive heart failure. The dominant expression of PPAR δ in cardiomyocytes of the heart explains at least in part the reason for mice with cardiomyocyte-restricted PPAR δ knockout exhibit a devastating pathological consequence [19, 30]. Thus PPAR δ may play a crucial role in metabolic homeostasis in the heart by maintaining intracellular FA content and a high-level FA metabolism observed in normal adult hearts.

Crosstalk or compensatory expression between PPAR δ and the other PPAR isoforms has been identified in the heart [69]. Nonetheless, pharmacological or genetically PPAR δ overexpression or activation did not modify the expression of PPAR α and PPAR γ in the heart in both physiological and pathological states [41, 69]. PPAR α and PPAR δ both regulate FA utilization in cardiomyocytes in an interdependent manner. Deletion of either of them from cardiomyocytes does not affect other PPARs' effects on activating FA utilization gene expression [19, 30]. Interestingly, cardiomyocyte-restricted PPAR δ knockout, but not the conventional PPAR α knockout mice, exhibit neutral lipid accumulation in myocardium at basal state [19, 71], probably due to the mismatch of PPAR δ activated FAO and lipid uptake. Until further investigation on a cardiomyocyte-restricted PPAR α KO mouse line, it is impossible to draw a definitive conclusion on the differentiated roles of these two important transcription factors in the heart. Interestingly, a study on a mouse line with cardiac-specific overexpression of a wild-type PPAR δ results in increased cardiac glucose uptake and oxidation in concomitant upregulation of glucose transporter type 4 (GLUT4) and phosphofructokinase (PFK) genes [46]. However, it remains unknown if these two genes are direct targets of PPAR δ . A recent study demonstrates that PPAR δ improves exercise endurance by reducing glucose and increasing fat metabolism in skeletal muscle [26]. Therefore, it is possible that the effects of PPAR δ on cardiac glucose metabolism may be indirect. Further studies will be warranted to clarify the direct effect of PPAR δ on myocardial glucose metabolism.

3.2. PPARS regulates mitochondrial antioxidant defense

The normal function and structure of the heart rely heavily on the efficiency of energy supply, which is dependent on the redox homeostasis of mitochondria, the power plant of cells. Mitochondrial redox dysfunction will result in energy deficiency and excessive production of reactive oxygen species (ROS) in cardiomyocytes, impairing cardiac function and structure. Therefore, a better understanding of the mitochondrial redox regulation in the heart may yield novel therapeutic strategies for treating cardiac disorders.

In adult mouse hearts with induced PPAR δ knockout, both mitochondrial biogenesis and cardiac antioxidant defense are impaired [40]. Specifically, PPAR δ knockout from adult heart leads to oxidative damages with an impaired cardiac expression of superoxide dismutase 1 (SOD1) and SOD2 [40]. PPAR α null mice with additional PPAR δ knockout from the heart showed similar results [69]. Both the transcript and protein expression of SOD1 and SOD2 was repressed in PPAR δ , but not PPAR α deficient hearts [69]. No endogenous antioxidants are affected at the basal condition in the PPAR α KO heart, supporting a notion that differential regulation of antioxidant expression in the heart may be the key factor contributing to the phenotypic changes in PPAR δ deficient hearts. The effects of PPAR δ activation in regulating antioxidant defense in the heart have also been validated in mouse models with cardiomyocyte-restricted overexpression of a constitutively active PPAR δ . With an enhanced antioxidant defense, the PPAR δ activating mice showed less oxidative injuries in the heart and stronger cardiac performance even under a left ventricular pressure overload condition [41]. Another study showed that activation of PPAR δ protects cardiomyocytes from oxidative stress-induced apoptosis by suppressing generation of reactive oxygen/nitrogen species and expression of matrix metalloproteinases [72]. The impaired expression of antioxidants should contribute to the cardiac dysfunction in the PPAR δ deficient heart [30] and activation of PPAR δ protects the heart at least partly via upregulating antioxidant levels in the heart under pressure overload condition [40, 41]. Other than the direct transcriptional regulation, the effect of PPAR δ in myocardial lipid metabolism and glucose utilization [46] at various metabolic states in different pathological development stages may contribute to the antioxidant defense and myocardial protection of PPAR δ ligand treatment and transgenic PPAR δ overexpression [40, 73].

Taken together, the above investigations support mostly a protective role of PPAR δ in regulating transcriptional expression of multiple endogenous anti-oxidants, protecting the heart under both physiological and pathological conditions.

3.3. PPARδ exerts anti-inflammation effects on the heart

Chronic inflammation is involved in the pathological development of myocardial ischemia/reperfusion injury [74, 75], pathological cardiac hypertrophy and congestive heart failure [76, 77]. Although remaining invalidated in patients, anti-inflammation to specific inflammation pathways remains a potential therapeutic target for the treatment of cardiac injuries and heart failure [78, 79].

The anti-inflammatory role of PPAR α and PPAR γ [80–82], later PPAR δ [83], has been documented in the last decade. The anti-inflammatory effects of PPARs in the cardiovascular system have been extensively investigated, especially in their anti-inflammatory effects on macrophage foam-cell formation and atheroscleosis [84, 85]. PPAR δ activation blocks lipid-induced inflammatory pathways in mouse heart and human cardiac cells [86]. PPAR δ represses transforming necrosis factor- α (TNF- α) in cultured cardiomyocytes via regulating the nuclear factor- κ B (NF- κ B) signaling pathway. We showed that a PPAR δ -selective ligand, GW0742, inhibits the lipopolysaccharide (LPS)-induced TNF- α production in cultured cardiomyocytes [87]. All these investigations strongly support that PPAR δ exerts significant anti-inflammatory effects in the heart.

Several studies established a consistent mechanism for the anti-inflammatory effects of PPAR δ by its direct or indirect interaction with NF- κ B signaling [42]. An early study has shown that unliganded PPAR δ sequesters the transcriptional repressor protein B cell lymphoma- 6 (Bcl6) and prevents it from binding to the response elements in the promoter regions of its target genes [88]. Bcl6 is released from PPAR δ and inhibits inflammatory signals [88–90]. PPAR δ can interact with other transcription factors such as the p65 subunit of NF– κ B, preventing NF– κ B dependent transcription [42]. Furthermore, the roles of PPAR δ in curbing palmitate-induced endoplasmic reticulum stress and inducing autophagy [91] may also contribute to its anti-inflammatory effects in cardiomyocytes.

Despite the well-recognized anti-inflammatory effects of PPAR δ in the heart, further studies are required to translate these findings into clinical treatment of heart disease. The main issue is still related to the potential side effects of the existed PPAR δ -selective ligands. On the other hand, the continuous discovery of naturally occurring ligands that could at least partially activating PPAR δ may help identify compounds that have less side effect yet process PPAR δ activation effects to achieve the therapeutic goal.

3.4. Potential adverse effects of PPARS in the heart

Although most investigations support the beneficial effects of PPAR δ on the heart, potentially detrimental effects of PPAR δ have been reported. In cultured H9C2 cells, a cell line of myoblasts from rat embryonic hearts with few features of cardiomyocytes, pharmacological blocking of PPAR δ signaling mitigates the cytotoxicity of docosahexaenoic acid (DHA), a key compound found in fish oil [92]. However, further *in vivo* studies are essential to validate this *in vitro* finding. Another study shows that PPAR δ ligands (GW0742 or GW501516) treatment in mice leads to rapid development of cardiac hypertrophy and angiogenesis, presumably via direct activation of calcineurin [93]. However, we did not see any cardiac hypertrophic effects of GW0742 with the same dose and subcutaneous injection. The reason for the discrepant cardiac effects of

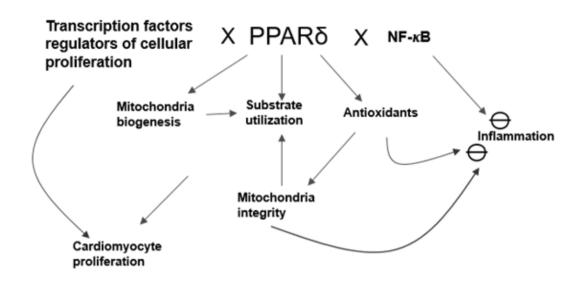
systemic treatment of GW0742 is not clear, although we cannot rule out a transient reversible cardiac growth induced by the treatment with GW0742 in our study [94]. The same group went on to demonstrate that mice with induced vascular-specific overexpression of PPAR δ show rapid cardiac hypertrophy [95]. Furthermore, the vascular overexpression of PPAR δ does not protect against chronic ischemic injury of the heart [95]. However, this study does not clarify if systemic treatment of GW0742 triggers cardiac hypertrophy by its effects on the vasculature. Therefore, further studies on the PPAR δ effects by assessing different cell types in the heart is needed in evaluating the potential clinical use of PPAR δ ligands for the treatment of heart diseases.

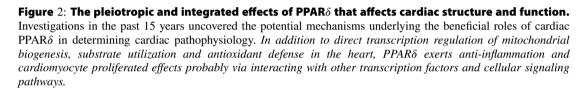
In summary, the importance of PPAR δ in cardiac pathophysiology is multi-faceted through direct transcriptional activation and secondary modulation of other transcriptional and cellular signaling pathways. For example, the PPAR δ interaction with NF-kB attributes to its antiinflammatory effects. It is also likely that the transcriptional effects of PPAR δ on FA utilization, mitochondrial biogenesis and antioxidant defense are further integrating with other biological effects such as inflammatory processes and cell proliferation (Figure 2). We recently uncovered that PPAR δ activation facilitates the cell cycle re-entry of adult cardiomyocyte [94]. Since adult cardiomyocytes are mostly terminally differentiated, enabling terminally differentiated cardiomyocyte re-entering cell cycle remains a major challenge [96–98]. Our drug screen identified carbacyclin, a PPAR δ activator, which induces cardiomyocyte proliferation via a PPAR δ /PDK1/p38/Akt/GSK3 β / β -catenin-pathway. Activating PPAR δ signaling after myocardial infarction (MI) induces cardiomyocyte cell cycle activity and improves scarring as well as cardiac function [94]. PPAR δ deletion from the heart reduces the effect of PPAR δ -selective agonist, GW0742, on adult cardiomyocyte proliferation in mice [94]. It is likely that potential interaction of PPAR δ with other transcriptional factors enables cardiomyocyte proliferation in the adult heart. Future investigations should focus on how PPAR δ interacts with other transcription factors/signaling pathways to exert its biological functions. Further studies should also target to exploit PPAR δ 's many potentially beneficial effects for the treatment of common cardiac disorders.

Upstream regulation of PPARδ

Several previous studies demonstrate that cardiac PPAR δ is downregulated in various pathological states [34, 35, 99]. The mechanisms underlying the downregulation of cardiac PPAR δ expression and activity is intriguing but remains elusive.

Nuclear receptor coactivator 6 (NCOA6) is a transcription coregulator. Mouse models of NCOA6 dysfunction develop severe dilated cardiomyopathy with impaired mitochondrial function and reduced activity of PPAR δ [100]. It is not clear if the down-regulation of cardiac PPAR δ is part of the pathological consequence or the co-regulator acts through an unknown mechanism that repressing the expression of PPAR δ . An exact mechanism will require further investigations. PPAR δ is modulated by epigenetic modification. Specific microRNAs repress the expression of cardiac PPAR δ . For example, the hypoxia-inducible microRNA cluster miR-199a-214 represses approximately PPAR δ target gene expression in the heart and impairs mitochondrial FAO [36, 70]. A recent study shows that the circadian repressors (crytochrome-1) CRY1 and CRY2 function as co-repressors for PPAR δ , which represses a distinct subset of PPAR δ





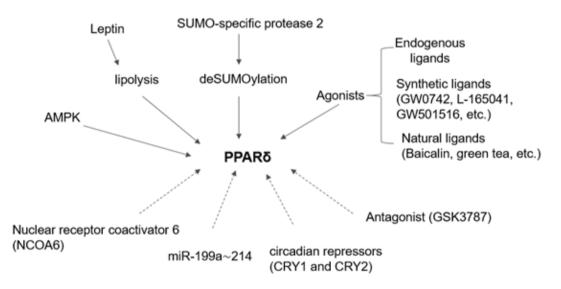


Figure 3: **Upstream regulation of PPAR** δ **in the heart.** The cardiac expression and activity of PPAR δ can be elevated by various PPAR δ agonists (endogenous, synthetic and natural ligands), post-translational modification (deSUMOylation), leptin and AMPK (AMP-activated protein kinase). In contrast, PPAR δ antagonists, microRNA (miR-199a-214) and nuclear receptor coregulator (Nuclear receptor coactivator 6). Arrows with solid lines: activation; Arrows with dash lines: inhibition.

target genes in skeletal muscle [101]. However, further study will be needed to determine if they exert similar PPAR δ repression effect in the heart.

A recent study revealed that leptin administered via intracerebroventricular enhances cardiac FAO by activating PPAR δ in the heart [102]. Various lipid species from the increased lipolysis by leptin down-stream signaling in the heart may serve as endogenous ligands that activate

cardiac PPAR δ by the central leptin. PPAR δ can be deactivated by post-translational modification such as SUMOylation. Small ubiquitin-like modifier (SUMO)-specific proteases (SENPs) that reverse protein modification by deSUMOylation of PPAR δ and PPAR γ to enhance FA metabolism in cultured C2C12 myotubes [103]. AMP-activated protein kinase (AMPK) is wellrecognized cellular energy-sensing enzyme complex [104, 105]. PPAR δ agonist GW501516 activates AMPK and stimulates glucose uptake in skeletal muscle [106, 107]. AMPK may interact with PPAR δ to exert their gene regulation function with unknown mechanisms [108]. Given the well-established beneficial effects of AMPK activation in the heart (see review [109]) and similarity of skeletal muscle and heart muscle, it is plausible that similar AMPK activation effects of PPAR δ activation may also apply to the heart. However, it remains obscure how AMPK is activated by PPAR δ , which presumably leads to increase energy metabolism. Further investigations are warranted. PPAR δ can also be activated by an array of long-chain FAs and prostaglandins and other metabolites. However, the specificity and potency of these endogenous ligands appear to be relatively modest. Several selective PPAR δ agonists have been developed and well documented, such as GW0742, GW501516, and L165041 [110-113]. Synthetic PPAR δ antagonists, such as GSK-3787, are also available [114]. Several clinically used therapeutic compounds have been reported to be able to enhance the activity of PPAR δ in addition to their specific therapeutic targets. Ramipril is an angiotensin-converting enzyme inhibitor, used to treat high blood pressure and congestive heart failure [115]. Interestingly, Ramipril can be normalized the decreased PPAR δ and PPAR γ mRNA in animals treated with the anti-cancer drug, daunorubicin [58]. The cholesterol-lowering drug, Atorvastatin, has also been reported to upregulate the expression of PPAR δ in Angiotensin II-induced hypertrophic cardiomyocytes *in vitro* [34]. Another study showed that digoxin could upregulate cardiac PPAR δ followed by improving lipid metabolism in the heart of diabetic rats [37]. Therefore, it appears that many currently used drugs may also exert PPAR δ activation effects, in addition to those well-designed and characterized PPAR δ -selective ligands. However, the underpinning mechanisms for their PPAR δ activation effects need further investigations.

Besides the endogenous and synthetic PPAR δ ligands, many naturally occurred compounds from plants have been shown to either act as PPAR δ ligands or PPAR δ enhancers. For example, green tea extract selectively activates PPAR δ in cultured cardiomyocytes [116, 117]. A recent study demonstrates that physiological levels of caffeine enhance cell metabolism at least partially via PPAR δ [118]. Although the *in vivo* effects of green tea and caffeine on the adult heart need further studies, the clinical implication in the tea and coffee drinking population is intriguing. Further study is required to determine if drinking tea or coffee may activate cardiac PPAR δ and may exert beneficial effects as those we shown in transgenic mice with heightening cardiomyocyte-restricted PPAR δ expression [40]. Baicalin is a natural flavone found in the extract of the rhizome of the perennial herb Scutellaria baicalensis, known as huangqin in China and is one of the most used herbs in Traditional Chinese Medicine for thousands of years [119]. Zhang et al. showed that baicalin restores cardiac PPAR δ expression in vivo and in vitro in cultured cardiomyocytes [35]. Abscistic acid, a phytohormone, is among those newly identified PPAR δ activators [120]. The cyclohexenyl chalcone panduratin A, isolated from *Boesenbergia* pandurata rhizomes, was shown to be a natural AMPK stimulator, with consequent activation of PPAR α/δ [121]. Therapeutic potential of panduratin A, an LKB1-dependent AMP-activated protein kinase stimulator, is shown to activate PPAR α/δ and can be an effective anti-obesity treatment [121]. Vaticanol C is reported to be a complex resveratrol tetramer activates PPAR α

and PPAR δ , but not PPAR γ both *in vitro* and *in vivo* [122]. Ginsenoside Rh2 treatment in type1-like diabetic rats can enhance cardiac expression of PPAR δ and repress cardiac fibrosis [123]. The list of new identified natural activators of PPAR is still expanding. However, it remains unclear how we may use these potential natural PPAR δ activators for prevention and alternative treatments of cardiac disorders. These studies do not provide potential mechanisms underlying the upregulation of cardiac PPAR δ . At present, studies are still absent to further determine which of the two PPAR subtypes (α and δ) exert the cardiac protective effects of these compounds. In systemic treatment, the cardiac PPAR δ activators could have been due to their effects on other systems, eg., PPAR δ exerts antihypertensive effects [124]. While the potential anti-hypertensive effect of activating PPAR δ post additional beneficial effect when treating heart failure patients with hypertension, other systemic effects of PPAR δ remain major confounding factors preventing the potential application. Therefore, it appears that the understanding of the molecular mechanisms for the specific changes in cardiac expression and activation of PPAR δ are surprisingly scarce. Further investigations in the regulation of cardiac PPAR δ may yield crucial insights into translatable therapies for treating heart disease.

5. Future Perspectives and Conclusion

As ligand-activated transcription factors, the PPARs represent an attractive target for the development of therapeutic agents. Synthetic PPAR δ agonists such as GW0742, GW501516, and L165041 [110–113] have been developed and even went on to early clinical trials. Although controversial, the main concern remains to be its potential tumorigenesis effects (see review [125]). The systemic use of PPAR δ is undoubtedly problematic because its seemingly differential effects on different cell types and tissues, which is evident by numerous studies using global and tissue-specific PPAR δ transgenic and gene-targeting mice [30, 41, 69, 126–128]. On-going efforts on designed-synthesized tissue-specific PPAR δ -selective ligands [129, 130] may give hope for targeting cardiac PPAR δ for the treatment of metabolic derangement and mitochondrial dysfunction in cardiac pathological states.

Despite the identification of numerous natural ligands for PPARs, a major challenge is the lack of in-depth studies on most of them. New approaches have been proposed using the cellular and biophysical pipeline for the screening of PPAR δ -selective ligands to avoid false positives [131]. Another alternative approach aims at further characterizing PPAR δ targets to delineate relevant downstream regulatory pathways to avoid the unwanted effects.

Acknowledements

This work was supported by grants from the National Natural Science Foundation of China (81500312 and 81570366 to Q. L.), American Diabetes Association (1-17-IBS-184 to Q.Y.), NIH (R01HL13533 and R03AG055899 to Q.Y.).

Competing Interests

The authors declare no competing interests.

References

- K. Schoonjans, G. Martin, B. Staels, and J. Auwerx, "Peroxisome proliferator-activated receptors, orphans with ligands and functions," *Current Opinion in Lipidology*, vol. 8, no. 3, pp. 159–166, 1997.
- [2] A. Schmidt, N. Endo, S. J. Rutledge, R. Vogel, D. Shinar, and G. A. Rodan, "Identification of a new member of the steroid hormone receptor superfamily that is activated by a peroxisome proliferator and fatty acids," *Molecular Endocrinology*, vol. 6, no. 10, pp. 1634–1641, 1992.
- [3] G. Krey, H. Keller, A. Mahfoudi et al., "Xenopus peroxisome proliferator activated receptors: Genomic organization, response element recognition, heterodimer formation with retinoid X receptor and activation by fatty acids," *The Journal of Steroid Biochemistry and Molecular Biology*, vol. 47, no. 1-6, pp. 65–73, 1993.
- [4] W. R. Oliver Jr., J. L. Shenk, M. R. Snaith et al., "A selective peroxisome proliferator-activated receptor δ agonist promotes reverse cholesterol transport," *Proceedings of the National Acadamy of Sciences of the United States* of America, vol. 98, no. 9, pp. 5306–5311, 2001.
- [5] U. Dressel, T. L. Allen, J. B. Pippal, P. R. Rohde, P. Lau, and G. E. O. Muscat, "The peroxisome proliferatoractivated receptor β/δ agonist, GW501516, regulates the expression of genes involved in lipid catabolism and energy uncoupling in skeletal muscle cells," *Molecular Endocrinology*, vol. 17, no. 12, pp. 2477–2493, 2003.
- [6] K. Iwaisako, M. Haimerl, Y.-H. Paik et al., "Protection from liver fibrosis by a peroxisome proliferator-activated receptor δ agonist," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 109, no. 21, pp. E1369–E1376, 2012.
- [7] D. Jones, P. F. Boudes, M. G. Swain et al., "Seladelpar (MBX-8025), a selective PPAR-δ agonist, in patients with primary biliary cholangitis with an inadequate response to ursodeoxycholic acid: a double-blind, randomised, placebo-controlled, phase 2, proof-of-concept study," *The Lancet Gastroenterology and Hepatology*, vol. 2, no. 10, pp. 716–726, 2017.
- [8] D. P. Kelly, "PPARs of the heart: Three is a crowd," Circulation Research, vol. 92, no. 5, pp. 482–484, 2003.
- [9] Y. Ding, KD. Yang, and Q. Yang, "The role of, pp. ardelta-ardeltsignaling in the cardiovascular system. Prog Mol Biol Transl Sci," in *The role of*, pp. 121–451, 121, 451-473, 2014.
- [10] T. Kim, "Peroxisome-proliferator-activated receptors regulate redox signaling in the cardiovascular system," *World Journal of Cardiology*, vol. 5, no. 6, p. 164, 2013.
- [11] L. Han, W.-J. Shen, S. Bittner, F. B. Kraemer, and S. Azhar, "PPARs: Regulators of metabolism and as therapeutic targets in cardiovascular disease. Part II: PPAR-β/δ and PPAR-γ," *Future Cardiology*, vol. 13, no. 3, pp. 279–296, 2017.
- [12] F. Djouadi, J. Brandt, C. Weinheimer, T. Leone, F. Gonzalez, and D. Kelly, "The role of the peroxisome proliferator-activated receptor α (PPARα) in the control of cardiac lipid metabolism," *Prostaglandins*, *Leukotrienes and Essential Fatty Acids*, vol. 60, no. 5-6, pp. 339–343, 1999.
- [13] P. M. Barger, J. M. Brandt, T. C. Leone, C. J. Weinheimer, and D. P. Kelly, "Deactivation of peroxisome proliferator-activated receptor-alpha during cardiac hypertrophic growth," *The Journal of Clinical Investigation*, vol. 105, no. 12, pp. 1723–1730, 2000.
- [14] S. Z. Duan, C. Y. Ivashchenko, M. W. Russell, D. S. Milstone, and R. M. Mortensen, "Cardiomyocyte-specific knockout and agonist of peroxisome proliferator-activated receptor-gamma both induce cardiac hypertrophy in mice," *Circulation Research*, vol. 97, no. 4, pp. 372–379, 2005.
- [15] G. Ding, M. Fu, Q. Qin et al., "Cardiac peroxisome proliferator-activated receptor gamma is essential in protecting cardiomyocytes from oxidative damage," *Cardiovascular Research*, vol. 76, no. 2, pp. 269–279, 2007.
- [16] J. Luo, S. Wu, J. Liu et al., "Conditional PPARγ knockout from cardiomyocytes of adult mice impairs myocardial fatty acid utilization and cardiac function," *American Journal of Translational Research*, vol. 3, no. 1, pp. 61–72, 2010.
- [17] N.-H. Son, S. Yu, J. Tuinei et al., "PPARγ-induced cardiolipotoxicity in mice is ameliorated by PPARα deficiency despite increases in fatty acid oxidation," *The Journal of Clinical Investigation*, vol. 120, no. 10, pp. 3443–3454, 2010.
- [18] A. J. Gilde, K. A. van der Lee, and P. H. Willemsen, "Peroxisome proliferator-activated receptor (PPAR) alpha and PPARbeta/delta, but not PPARgamma, modulate the expression of genes involved in cardiac lipid metabolism," *Circulation Research*, vol. 92, no. 5, pp. 518–524, 2003.
- [19] L. Cheng, G. Ding, Q. Qin et al., "Peroxisome proliferator-activated receptor δ activates fatty acid oxidation in cultured neonatal and adult cardiomyocytes," *Biochemical and Biophysical Research Communications*, vol. 313, no. 2, pp. 277–286, 2004.
- [20] Y. Barak, D. Liao, W. He et al., "Effects of peroxisome proliferator-activated receptor δ on placentation, adiposity, and colorectal cancer," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 99, no. 1, pp. 303–308, 2002.
- [21] S. A. Kliewer, B. M. Forman, B. Blumberg et al., "Differential expression and activation of a family of murine peroxisome proliferator-activated receptors," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 91, no. 15, pp. 7355–7359, 1994.

- [22] O. Braissant, F. Foufelle, C. Scotto, M. Dauça, and W. Wahli, "Differential expression of peroxisome proliferatoractivated receptors (PPARs): tissue distribution of PPAR-α, -β, and -γ in the adult rat," *Endocrinology*, vol. 137, no. 1, pp. 354–366, 1996.
- [23] A. R. Angione, C. Jiang, D. Pan, Y.-X. Wang, and S. Kuang, "PPARδ regulates satellite cell proliferation and skeletal muscle regeneration," *Skeletal Muscle*, vol. 1, no. 1, 2011.
- [24] S. Beyaz, M. D. Mana, J. Roper et al., "High-fat diet enhances stemness and tumorigenicity of intestinal progenitors," *Nature*, vol. 531, no. 7592, pp. 53–58, 2016.
- [25] T. Degenhardt, A. Saramäki, M. Malinen et al., "Three Members of the Human Pyruvate Dehydrogenase Kinase Gene Family Are Direct Targets of the Peroxisome Proliferator-activated Receptor β/δ," *Journal of Molecular Biology*, vol. 372, no. 2, pp. 341–355, 2007.
- [26] W. Fan, W. Waizenegger, C. S. Lin et al., "PPARδ Promotes Running Endurance by Preserving Glucose," Cell Metabolism, vol. 25, no. 5, pp. 1186–1193.e4, 2017.
- [27] K. Ito, A. Carracedo, D. Weiss et al., "A PML-PPAR-δ pathway for fatty acid oxidation regulates hematopoietic stem cell maintenance," *Nature Medicine*, vol. 18, no. 9, pp. 1350–1358, 2012.
- [28] H. E. Kocalis, M. K. Turney, R. L. Printz et al., "Neuron-specific deletion of peroxisome proliferator-activated receptor delta (PPAR δ) in mice leads to increased susceptibility to diet-induced obesity," *PLoS ONE*, vol. 7, no. 8, 2012.
- [29] K. Kang, S. M. Reilly, V. Karabacak et al., "Adipocyte-derived Th2 cytokines and myeloid PPARd regulate macrophage polarization and insulin sensitivity," *Cell Metabolism*, vol. 7, no. 6, pp. 485–495, 2008.
- [30] L. Cheng, G. Ding, Q. Qin et al., "Cardiomyocyte-restricted peroxisome proliferator-activated receptor-δ deletion perturbs myocardial fatty acid oxidation and leads to cardiomyopathy," *Nature Medicine*, vol. 10, no. 11, pp. 1245–1250, 2004.
- [31] W. C. Stanley, F. A. Recchia, and G. D. Lopaschuk, "Myocardial substrate metabolism in the normal and failing heart," *Physiological Reviews*, vol. 85, no. 3, pp. 1093–1129, 2005.
- [32] V. Lionetti, W. C. Stanley, and F. A. Recchia, "Modulating fatty acid oxidation in heart failure," *Cardiovascular Research*, vol. 90, no. 2, pp. 202–209, 2011.
- [33] M. Vázquez-Carrera, "Unraveling the effects of PPARβ/δ on insulin resistance and cardiovascular disease," *Trends in Endocrinology & Metabolism*, vol. 27, no. 5, pp. 319–334, 2016.
- [34] L. Sheng, X. Yang, P. Ye, Y.-X. Liu, and C.-G. Han, "Effect of Atorvastatin on Expression of Peroxisome Proliferator-activated Receptor Beta/delta in Angiotensin II-induced Hypertrophic Myocardial Cells in Vitro," *Chinese Medical Sciences Journal*, vol. 30, no. 4, pp. 245–251, 2015.
- [35] Y. Zhang, P. Liao, M. Zhu et al., "Baicalin Attenuates Cardiac Dysfunction and Myocardial Remodeling in a Chronic Pressure-Overload Mice Model," *Cellular Physiology and Biochemistry*, vol. 41, no. 3, pp. 849–864, 2017.
- [36] H. El Azzouzi, S. Leptidis, E. Dirkx et al., "The hypoxia-inducible microRNA cluster miR-199a approximately 214 targets myocardial PPARdelta and impairs mitochondrial fatty acid oxidation," *Cell Metabolism*, vol. 18, no. 3, pp. 341–354, 2013.
- [37] Z. Chen, B. Yu, L. Chen, and J. Cheng, "Increase of Peroxisome Proliferator-activated Receptor δ (PPAR δ) by Digoxin to Improve Lipid Metabolism in the Heart of Diabetic Rats," *Hormone and Metabolic Research*, vol. 45, no. 05, pp. 364–371, 2013.
- [38] S. Lee, G. Go, M. Yoo et al., "Peroxisome proliferator-activated receptor β/δ (PPARβ/δ) activates promyogenic signaling pathways, thereby promoting myoblast differentiation," *Biochemical and Biophysical Research Communications*, vol. 470, no. 1, pp. 157–162, 2016.
- [39] A. Kapoor, M. Collino, S. Castiglia, R. Fantozzi, and C. Thiemermann, "Activation of peroxisome proliferatoractivated receptor-β/δ attenuates myocardial ischemia/reperfusion injury in the rat," *Shock*, vol. 34, no. 2, pp. 117–124, 2010.
- [40] P. Wang, J. Liu, Y. Li et al., "Peroxisome proliferator-activated receptor δ is an essential transcriptional regulator for mitochondrial protection and biogenesis in adult heart," *Circulation Research*, vol. 106, no. 5, pp. 911–919, 2010.
- [41] J. Liu, P. Wang, J. Luo et al., "Peroxisome proliferator-activated receptor β/δ activation in adult hearts facilitates mitochondrial function and cardiac performance under pressure-overload condition," *Hypertension*, vol. 57, no. 2, pp. 223–230, 2011.
- [42] A. Planavila, R. Rodríguez-Calvo, M. Jové et al., "Peroxisome proliferator-activated receptor β/δ activation inhibits hypertrophy in neonatal rat cardiomyocytes," *Cardiovascular Research*, vol. 65, no. 4, pp. 832–841, 2005.
- [43] C. Pellieux, C. Montessuit, I. Papageorgiou, and R. Lerch, "Inactivation of peroxisome proliferator-activated receptor isoforms α, β/δ, and γ mediate distinct facets of hypertrophic transformation of adult cardiac myocytes," *Pflügers Archiv - European Journal of Physiology*, vol. 455, no. 3, pp. 443–454, 2007.
- [44] B. Kojonazarov, H. Luitel, A. Sydykov et al., "The peroxisome proliferator-activated receptor β/δ agonist GW0742 has direct protective effects on right heart hypertrophy," *Pulmonary Circulation*, vol. 3, no. 4, pp. 926–935, 2013.

- [45] J. I. Randriamboavonjy, G. Loirand, N. Vaillant et al., "Cardiac protective effects of moringa oleifera seeds in spontaneous hypertensive rats," *American Journal of Hypertension*, vol. 29, no. 7, pp. 873–881, 2016.
- [46] E. M. Burkart, N. Sambandam, X. Han et al., "Nuclear receptors PPARβ/δ and PPARα direct distinct metabolic regulatory programs in the mouse heart," *The Journal of Clinical Investigation*, vol. 117, no. 12, pp. 3930–3939, 2007.
- [47] A. Magadum, Y. Ding, L. He et al., "Live cell screening platform identifies PPAR δ as a regulator of cardiomyocyte proliferation and cardiac repair," *Cell Research*, vol. 27, no. 8, pp. 1002–1019, 2017.
- [48] J. R. Park, J. H. Ahn, M. H. Jung et al., "Effects of Peroxisome Proliferator-Activated Receptor-δ Agonist on Cardiac Healing after Myocardial Infarction," *PLoS ONE*, vol. 11, no. 2, p. e0148510, 2016.
- [49] P. Altieri, P. Spallarossa, C. Barisione et al., "Inhibition of Doxorubicin-Induced Senescence by PPARδ Activation Agonists in Cardiac Muscle Cells: Cooperation between PPARδ and Bcl6," *PLoS ONE*, vol. 7, no. 9, 2012.
- [50] Zhih-Cherng Chen, Li-Jen Chen, and Juei-Tang Cheng, "Doxorubicin-Induced Cardiac Toxicity Is Mediated by Lowering of Peroxisome Proliferator-Activated Receptor δ Expression in Rats," *PPAR Research*, vol. 2013, Article ID 456042, 8 pages, 2013.
- [51] W.-T. Chang, J. Li, H.-H. Haung et al., "Baicalein protects against doxorubicin-induced cardiotoxicity by attenuation of mitochondrial oxidant injury and JNK activation," *Journal of Cellular Biochemistry*, vol. 112, no. 10, pp. 2873–2881, 2011.
- [52] M. Bayeva, K. T. Sawicki, and H. Ardehali, "Taking diabetes to heart—deregulation of myocardial lipid metabolism in diabetic cardiomyopathy," *Journal of the American Heart Association*, vol. 2, no. 6, Article ID e000433, 2013.
- [53] S. Boudina and E. D. Abel, "Diabetic cardiomyopathy revisited," *Circulation*, vol. 115, no. 25, pp. 3213–3223, 2007.
- [54] J. D. Schilling, "The Mitochondria in Diabetic Heart Failure: From Pathogenesis to Therapeutic Promise," *Antioxidants & Redox Signaling*, vol. 22, no. 17, pp. 1515–1526, 2015.
- [55] A. Georgiadi, L. Lichtenstein, T. Degenhardt et al., "Induction of cardiac angpt14 by dietary fatty acids is mediated by peroxisome proliferator-activated receptor β/δ and protects against fatty acid-induced oxidative stress," *Circulation Research*, vol. 106, no. 11, pp. 1712–1721, 2010.
- [56] T.-I. Lee, Y.-H. Kao, W.-C. Tsai, C.-C. Chung, Y.-C. Chen, and Y.-J. Chen, "HDAC inhibition modulates cardiac PPARs and fatty acid metabolism in diabetic cardiomyopathy," *PPAR Research*, vol. 2016, 2016.
- [57] Cheng-Chia Tsai, Paul Chan, Li-Jen Chen, Chen Kuei Chang, Zhongmin Liu, and Jia-Wei Lin, "Merit of Ginseng in the Treatment of Heart Failure in Type 1-Like Diabetic Rats," *BioMed Research International*, vol. 2014, Article ID 484161, 8 pages, 2014.
- [58] H. Cernecka, G. Doka, J. Srankova et al., "Ramipril restores PPAR β/δ and PPAR γ expressions and reduces cardiac NADPH oxidase but fails to restore cardiac function and accompanied myosin heavy chain ratio shift in severe anthracycline-induced cardiomyopathy in rat," *European Journal of Pharmacology*, vol. 791, pp. 244–253, 2016.
- [59] B. N. Finck, J. J. Lehman, T. C. Leone et al., "The cardiac phenotype induced by PPARα overexpression mimics that caused by diabetes mellitus," *The Journal of Clinical Investigation*, vol. 109, no. 1, pp. 121–130, 2002.
- [60] M. E. Young, F. A. Laws, G. W. Goodwin, and H. Taegtmeyer, "Reactivation of peroxisome proliferator-activated receptor alpha is associated with contractile dysfunction in hypertrophied rat heart," *The Journal of Biological Chemistry*, vol. 276, no. 48, pp. 44390–44395, 2001.
- [61] J. Zou, K. Le, S. Xu et al., "Fenofibrate ameliorates cardiac hypertrophy by activation of peroxisome proliferatoractivated receptor-alpha partly via preventing p65-NFkappaB binding to NFATc4," *Molecular and Cellular Endocrinology*, vol. 370, no. 1-2, pp. 103–112, 2013.
- [62] V. H. Lam, L. Zhang, A. Huqi et al., "Activating PPARα Prevents Post-Ischemic Contractile Dysfunction in Hypertrophied Neonatal Hearts," *Circulation Research*, vol. 117, no. 1, pp. 41–51, 2015.
- [63] Si-Chi Xu, Zhen-Guo Ma, Wen-Ying Wei, Yu-Pei Yuan, and Qi-Zhu Tang, "Bezafibrate Attenuates Pressure Overload-Induced Cardiac Hypertrophy and Fibrosis," *PPAR Research*, vol. 2017, Article ID 5789714, 12 pages, 2017.
- [64] S. Rana, R. Datta, R. D. Chaudhuri, E. Chatterjee, M. Chawla-Sarkar, and S. Sarkar, "Nanotized PPARα overexpression targeted to hypertrophied myocardium improves cardiac function by attenuating the p53-GSK3β mediated mitochondrial death pathway," *Antioxidants & Redox Signaling*, 2018.
- [65] X. Palomer, E. Barroso, M. Zarei, G. Botteri, M. Vazquez-Carrera, and pp., "arbeta-arbetdelta and lipid metabolism in the heart," *Biochimica et Biophysica Acta*, pp. 1861–1569, 2016.
- [66] Wang-Soo Lee and Jaetaek Kim, "Peroxisome Proliferator-Activated Receptors and the Heart: Lessons from the Past and Future Directions," *PPAR Research*, vol. 2015, Article ID 271983, 18 pages, 2015.
- [67] Q. Yang and L. Cheng, "Molecular regulation of lipotoxicity in the heart," Drug Discovery Today: Disease Mechanisms, vol. 2, no. 1, pp. 101–107, 2005.

- [68] Q. Yang and Y. Li, "Roles of PPARs on regulating myocardial energy and lipid homeostasis," *Journal of Molecular Medicine*, vol. 85, no. 7, pp. 697–706, 2007.
- [69] Jian Liu, Peiyong Wang, Lan He et al., "Cardiomyocyte-Restricted Deletion of PPARβ/δ in PPARα-Null Mice Causes Impaired Mitochondrial Biogenesis and Defense, but No Further Depression of Myocardial Fatty Acid Oxidation," PPAR Research, vol. 2011, Article ID 372854, 13 pages, 2011.
- [70] S. Kuo, P. Ku, L. Chen, H. Niu, and J. Cheng, "Activation of Receptors δ (PPARδ) by Agonist (GW0742) may Enhance Lipid Metabolism in Heart both In Vivo and In Vitro," *Hormone and Metabolic Research*, vol. 45, no. 12, pp. 880–886, 2013.
- [71] K. Watanabe, H. Fujii, T. Takahashi et al., "Constitutive regulation of cardiac fatty acid metabolism through peroxisome proliferator-activated receptor α associated with age-dependent cardiac toxicity," *The Journal of Biological Chemistry*, vol. 275, no. 29, pp. 22293–22299, 2000.
- [72] E. Barlaka, A. Görbe, R. Gáspár, J. Pálóczi, P. Ferdinandy, and A. Lazou, "Activation of PPARβ/δ protects cardiac myocytes from oxidative stress-induced apoptosis by suppressing generation of reactive oxygen/nitrogen species and expression of matrix metalloproteinases," *Pharmacological Research*, vol. 95-96, pp. 102–110, 2015.
- [73] T. L. Yue, S. S. Nerurkar, W. Bao et al., "In vivo activation of peroxisome proliferator-activated receptor-δ protects the heart from ischemia/reperfusion injury in Zucker fatty rats," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 325, no. 2, pp. 466–474, 2008.
- [74] J. L. Mehta, W. W. Nichols, and P. Mehta, "Neutrophils as potential participants in acute myocardial ischemia: relevance to reperfusion," *Journal of the American College of Cardiology*, vol. 11, no. 6, pp. 1309–1316, 1988.
- [75] N. G. Frangogiannis, "The inflammatory response in myocardial injury, repair, and remodelling," *Nature Reviews Cardiology*, vol. 11, no. 5, pp. 255–265, 2014.
- [76] P. J. H. Smeets, B. E. J. Teunissen, A. Planavila et al., "Inflammatory pathways are activated during cardiomyocyte hypertrophy and attenuated by peroxisome proliferator-activated receptors PPARα and PPARδ," *The Journal of Biological Chemistry*, vol. 283, no. 43, pp. 29109–29118, 2008.
- [77] A. B. Cohen, M. D. Stevens, E. J. Miller et al., "Neutrophil-activating peptide-2 in patients with pulmonary edema from congestive heart failure or ARDS," *American Journal of Physiology-Lung Cellular and Molecular Physiology*, vol. 264, no. 5, pp. L490–L495, 1993.
- [78] S. Heymans, E. Hirsch, S. D. Anker et al., "Inflammation as a therapeutic target in heart failure? A scientific statement from the Translational Research Committee of the Heart Failure Association of the European Society of Cardiology," *European Journal of Heart Failure*, vol. 11, no. 2, pp. 119–129, 2009.
- [79] N. Suthahar, W. C. Meijers, H. H. W. Silljé, and R. A. de Boer, "From Inflammation to Fibrosis—Molecular and Cellular Mechanisms of Myocardial Tissue Remodelling and Perspectives on Differential Treatment Opportunities," *Current Heart Failure Reports*, vol. 14, no. 4, pp. 235–250, 2017.
- [80] S. M. Jackson, F. Parhami, X.-P. Xi et al., "Peroxisome proliferator-activated receptor activators target human endothelial cells to inhibit leukocyte-endothelial cell interaction," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 19, no. 9, pp. 2094–2104, 1999.
- [81] C. G. Su, X. Wen, S. T. Bailey et al., "A novel therapy for colitis utilizing PPAR-γ ligands to inhibit the epithelial inflammatory response," *The Journal of Clinical Investigation*, vol. 104, no. 4, pp. 383–389, 1999.
- [82] J.-C. Fruchart, P. Duriez, and B. Staels, "Peroxisome proliferator-activated receptor-alpha activators regulate genes governing lipoprotein metabolism, vascular inflammation and atherosclerosis," *Current Opinion in Lipidology*, vol. 10, no. 3, pp. 245–257, 1999.
- [83] N. S. Tan, L. Michalik, N. Noy et al., "Critical roles of PPARbeta/delta in keratinocyte response to inflammation," *Genes & Development*, vol. 15, no. 24, pp. 3263–3277, 2001.
- [84] A. C. Li, C. J. Binder, A. Gutierrez et al., "Differential inhibition of macrophage foam-cell formation and atherosclerosis in mice by PPAR α , β/δ , and γ ," *The Journal of Clinical Investigation*, vol. 114, no. 11, pp. 1564–1576, 2004.
- [85] Y. Liang, S. Lin, T. P. Beyer et al., "A liver X receptor and retinoid X receptor heterodimer mediates apolipoprotein E expression, secretion and cholesterol homeostasis in astrocytes," *Journal of Neurochemistry*, vol. 88, no. 3, pp. 623–634, 2004.
- [86] D. Álvarez-Guardia, X. Palomer, T. Coll et al., "PPARβ/δ activation blocks lipid-induced inflammatory pathways in mouse heart and human cardiac cells," *Biochimica et Biophysica Acta*, vol. 1811, no. 2, pp. 59–67, 2011.
- [87] G. Ding, L. Cheng, Q. Qin, S. Frontin, and Q. Yang, "PPARδ modulates lipopolysaccharide-induced TNFα inflammation signaling in cultured cardiomyocytes," *Journal of Molecular and Cellular Cardiology*, vol. 40, no. 6, pp. 821–828, 2006.
- [88] C. Lee, A. Chawla, N. Urbiztondo, D. Liao, W. A. Boisvert, and R. M. Evans, "Transcriptional repression of atherogenic inflammation: modulation by PPARδ," *Science*, vol. 302, no. 5644, pp. 453–457, 2003.
- [89] Y. Fan, Y. Wang, Z. Tang et al., "Suppression of pro-inflammatory adhesion molecules by PPAR-δ in human vascular endothelial cells," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 28, no. 2, pp. 315–321, 2008.

- [90] Y. Takata, J. Liu, F. Yin et al., "PPAR -mediated antiinflammatory mechanisms inhibit angiotensin II-accelerated atherosclerosis," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 105, no. 11, pp. 4277–4282, 2008.
- [91] X. Palomer, E. Capdevila-Busquets, G. Botteri et al., "PPARβ/δ attenuates palmitate-induced endoplasmic reticulum stress and induces autophagic markers in human cardiac cells," *International Journal of Cardiology*, vol. 174, no. 1, pp. 110–118, 2014.
- [92] V. Samokhvalov, I. Zlobine, K. L. Jamieson et al., "PPAR δ signaling mediates the cytotoxicity of DHA in H9c2 cells," *Toxicology Letters*, vol. 232, no. 1, pp. 10–20, 2015.
- [93] N. Wagner, C. Jehl-Piétri, P. Lopez et al., "Peroxisome proliferator-activated receptor β stimulation induces rapid cardiac growth and angiogenesis via direct activation of calcineurin," *Cardiovascular Research*, vol. 83, no. 1, pp. 61–71, 2009.
- [94] T. A. Ajith and T. G. Jayakumar, "Peroxisome proliferator-activated receptors in cardiac energy metabolism and cardiovascular disease," *Clinical and Experimental Pharmacology and Physiology*, vol. 43, no. 7, pp. 649–658, 2016.
- [95] K.-D. Wagner, A. Vukolic, D. Baudouy, J.-F. Michiels, and N. Wagner, "Inducible conditional vascular-specific overexpression of peroxisome proliferator-activated receptor beta/delta leads to rapid cardiac hypertrophy," *PPAR Research*, vol. 2016, 2016.
- [96] A. T. Akhmedov and J. Marín-García, "Myocardial regeneration of the failing heart," *Heart Failure Reviews*, vol. 18, no. 6, pp. 815–833, 2013.
- [97] K. E. Yutzey, "Cardiomyocyte proliferation," Circulation Research, vol. 120, no. 4, pp. 627–629, 2017.
- [98] X. Yuan and T. Braun, "Multimodal Regulation of Cardiac Myocyte Proliferation," *Circulation Research*, vol. 121, no. 3, pp. 293–309, 2017.
- [99] Z. Chen, K. S. Lee, L. Chen et al., "Cardiac Peroxisome Proliferator-Activated Receptor δ (PPAR δ) as a New Target for Increased Contractility without Altering Heart Rate," *PLoS ONE*, vol. 8, no. 5, p. e64229, 2013.
- [100] J.-I. Roh, C. Cheong, Y. Sung et al., "Perturbation of NCOA6 leads to dilated cardiomyopathy," *Cell Reports*, vol. 8, no. 4, pp. 991–998, 2014.
- [101] S. D. Jordan, A. Kriebs, M. Vaughan et al., "CRY1/2 Selectively Repress PPARδ and Limit Exercise Capacity," *Cell Metabolism*, vol. 26, no. 1, pp. 243–255.e6, 2017.
- [102] C. Mora, C. Pintado, B. Rubio et al., "Central leptin regulates heart lipid content by selectively increasing PPAR β/δ expression," Journal of Endocrinology, vol. 236, no. 1, pp. 43–56, 2018.
- [103] Y. D. Koo, J. W. Choi, M. Kim et al., "SUMO-Specific Protease 2 (SENP2) is an important regulator of fatty acid metabolism in skeletal muscle," *Diabetes*, vol. 64, no. 7, pp. 2420–2431, 2015.
- [104] Z. H. Beg, D. W. Allmann, and D. M. Gibson, "Modulation of 3-hydroxy-3-methylglutaryl coenzyme A reductase activity with cAMP and with protein fractions of rat liver cytosol," *Biochemical and Biophysical Research Communications*, vol. 54, no. 4, pp. 1362–1369, 1973.
- [105] D. Garcia and R. J. Shaw, "AMPK: Mechanisms of Cellular Energy Sensing and Restoration of Metabolic Balance," *Molecular Cell*, vol. 66, no. 6, pp. 789–800, 2017.
- [106] Z. Gan, E. M. Burkart-Hartman, D.-H. Han et al., "The nuclear receptor PPARβ/δ programs muscle glucose metabolism in cooperation with AMPK and MEF2," *Genes & Development*, vol. 25, no. 24, pp. 2619–2630, 2011.
- [107] L. Salvadó, E. Barroso, A. M. Gómez-Foix et al., "PPARβ/δ prevents endoplasmic reticulum stress-associated inflammation and insulin resistance in skeletal muscle cells through an AMPK-dependent mechanism," *Diabetologia*, vol. 57, no. 10, pp. 2126–2135, 2014.
- [108] M. Kemmerer, F. Finkernagel, M. F. Cavalcante et al., "AMP-activated protein kinase interacts with the peroxisome proliferator-activated receptor delta to induce genes affecting fatty acid oxidation in human macrophages," *PLoS ONE*, vol. 10, no. 6, Article ID e0130893, 2015.
- [109] T. T. Kim and J. R. Dyck, "Is AMPK the savior of the failing heart?" Trends in Endocrinology & Metabolism, vol. 26, no. 1, pp. 40–48, 2015.
- [110] J. Berger, M. D. Leibowitz, T. W. Doebber et al., "Novel peroxisome proliferator-activated receptor (PPAR) γ and PPAR δ ligands produce distinct biological effects," *The Journal of Biological Chemistry*, vol. 274, no. 10, pp. 6718–6725, 1999.
- [111] M. D. Leibowitz, C. Fiévet, N. Hennuyer et al., "Activation of PPARδ alters lipid metabolism in db/db mice," *FEBS Letters*, vol. 473, no. 3, pp. 333–336, 2000.
- [112] G. Wolf, "The Function of the Nuclear Receptor Peroxisome Proliferator-activated Receptor Delta in Energy Homeostasis," *Nutrition Reviews*, vol. 61, no. 11, pp. 387–390, 2003.
- [113] M. L. Sznaidman, C. D. Haffner, P. R. Maloney et al., "Novel selective small molecule agonists for peroxisome proliferator-activated receptor δ (PPAR δ) Synthesis and biological activity," *Bioorganic & Medicinal Chemistry Letters*, vol. 13, no. 9, pp. 1517–1521, 2003.

- [114] B. G. Shearer, R. W. Wiethe, A. Ashe et al., "Identification and characterization of 4-chloro-N-(2- [5-trifluoromethyl)- 2-pyridyl] sulfonyl ethyl)benzamide (GSK3787), a selective and irreversible peroxisome proliferator-activated receptor δ (PPARδ) antagonist," *Journal of Medicinal Chemistry*, vol. 53, no. 4, pp. 1857–1861, 2010.
- [115] L. Pilote, M. Abrahamowicz, M. Eisenberg, K. Humphries, H. Behlouli, and J. V. Tu, "Effect of different angiotensin-converting-enzyme inhibitors on mortality among elderly patients with congestive heart failure," *Canadian Medical Association Journal*, vol. 178, no. 10, pp. 1303–1311, 2008.
- [116] F. Danesi, M. di Nunzio, E. Boschetti, and A. Bordoni, "Green tea extract selectively activates peroxisome proliferator-activated receptor beta/delta in cultured cardiomyocytes," *British Journal of Nutrition*, vol. 101, no. 12, pp. 1736–1739, 2009.
- [117] S. Hrelia, A. Bordoni, C. Angeloni et al., "Green tea extracts can counteract the modification of fatty acid composition induced by doxorubicin in cultured cardiomyocytes," *Prostaglandins, Leukotrienes and Essential Fatty Acids*, vol. 66, no. 5-6, pp. 519–524, 2002.
- [118] J. K. Schnuck, L. M. Gould, H. A. Parry et al., "Metabolic effects of physiological levels of caffeine in myotubes," *Journal of Physiology and Biochemistry*, vol. 74, no. 1, pp. 35–45, 2018.
- [119] Y. Zhu, [the study on baicalin in china]. Zhongguo Zhong Xi Yi Jie He Za Zhi, 13, 567-569, 1993.
- [120] M. Mollashahi, M. Abbasnejad, and S. Esmaeili-Mahani, "Phytohormone abscisic acid elicits antinociceptive effects in rats through the activation of opioid and peroxisome proliferator-activated receptors β/δ ," *European Journal of Pharmacology*, vol. 832, pp. 75–80, 2018.
- [121] D. Kim, M. Lee, K. Jo, K. Lee, and J. Hwang, "Therapeutic potential of panduratin A, LKB1-dependent AMPactivated protein kinase stimulator, with activation of PPAR α/δ for the treatment of obesity," *Diabetes, Obesity* and Metabolism, vol. 13, no. 7, pp. 584–593, 2011.
- [122] T. Tsukamoto, R. Nakata, E. Tamura et al., "Vaticanol C, a resveratrol tetramer, activates PPARα and PPARβ/δ in vitro and in vivo," *Nutrition & Metabolism*, vol. 7, no. 1, p. 46, 2010.
- [123] S. Lo, C. Hsu, H. Niu, C. Niu, J. Cheng, and Z. Chen, "Ginsenoside Rh2 Improves Cardiac Fibrosis via PPARδ– STAT3 Signaling in Type 1-Like Diabetic Rats," *International Journal of Molecular Sciences*, vol. 18, no. 7, p. 1364, 2017.
- [124] M. Toral, M. Romero, F. Pérez-Vizcaíno, J. Duarte, and R. Jiménez, "Antihypertensive effects of peroxisome proliferator-activated receptor-β/δ activation," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 312, no. 2, pp. H189–H200, 2017.
- [125] R. Muller and pp. arbeta-arbetdelta in, "human cancer," Biochimie, pp. 136–90, 2017.
- [126] Y.-X. Wang, C.-H. Lee, S. Tiep et al., "Peroxisome-proliferator-activated receptor δ activates fat metabolism to prevent obesity," *Cell*, vol. 113, no. 2, pp. 159–170, 2003.
- [127] L. V. d'Uscio, T. He, A. V. R. Santhanam, L.-J. Tai, R. M. Evans, and Z. S. Katusic, "Mechanisms of vascular dysfunction in mice with endothelium-specific deletion of the PPAR-delta gene," *American Journal* of *Physiology-Heart and Circulatory Physiology*, vol. 306, no. 7, pp. H1001–H1010, 2014.
- [128] X. Zuo, Z. Peng, M. J. Moussalli et al., "Targeted genetic disruption of peroxisome proliferator-activated receptor-δ and colonic tumorigenesis," *Journal of the National Cancer Institute*, vol. 101, no. 10, pp. 762–767, 2009.
- [129] A. Bugge and D. Holst, "PPAR agonists, Could tissue targeting pave the way?" *Biochimie*, vol. 136, pp. 100–104, 2017.
- [130] V. G. Maltarollo, T. Kronenberger, B. Windshugel, C. Wrenger, G. H. Trossini, and K. M. Honorio, "Advances and Challenges in Drug Design of PPARδ Ligands," *Current Drug Targets*, vol. 19, no. 2, pp. 144–154, 2018.
- [131] Natália Bernardi Videira, Fernanda Aparecida Heleno Batista, Artur Torres Cordeiro, and Ana Carolina Migliorini Figueira, "Cellular and Biophysical Pipeline for the Screening of Peroxisome Proliferator-Activated Receptor Beta/Delta Agonists: Avoiding False Positives," *PPAR Research*, vol. 2018, Article ID 3681590, 14 pages, 2018.